National Laboratory for HIV Reference Services
National HIV and Retrovirology Laboratories
National Microbiology Laboratory
Public Health Agency of Canada

HTLV Serology Quality Assessment Program

Revised Summary for Panel HTLVSER 2021Apr19

2021Apr19 HTLV Serology Panel		
Panel Sample	**True Status**	**Labs Reporting Incorrect Status**
A	HTLV-II Ab Positive	
B	HTLV-I Ab Positive	
C	Negative	
D	HTLV-I Ab Positive	
E	Negative	

No incorrect results were observed for the 2021Apr19 test event.

In the original report, sample A (MP Diagnostic HTLV Blot 2.4 WB assay) is presented to have the p32 band as "1" and the p28 and p19 bands as "-". This is incorrect. The p32 band should be presented as "-" whereas the p28 and p19 bands should be "1" based on the results submitted by the participant. This revised report is reflective of the correction in Appendix 3, with changes highlighted in grey.

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Final Report for Panel HTLVSER 2021Apr19

Originally Issued 2021-July-19 and Revised 2021-August-13

Introduction

The NLHRS distributed the 2020Oct30 and 2021Apr19 panels on October 14, 2020. This final report is specific to the 2021Apr19 panel only and is publicly available; however, the identity of participants has not been disclosed. The deadline for results submission was April 19, 2021. The preliminary report was issued on May 5, 2021. The final report was amended due to transcriptional errors found in Appendix 3. The corrections are highlighted in grey.

Panel Samples, HTLV Test Kits, and Data Entry

- *Panel Composition*
  - The 2021Apr19 panel consisted of five samples: two HTLV negative (C, E), two HTLV-I positive (B, D), and one HTLV-II positive sample (A). Samples B and D were diluted 1 in 2 with defibrinated human plasma (Basematrix 53, Seracare Life Sciences). Testing and characterization by the NLHRS are presented in Appendix 1. Panels were sent to 17 participants including the NLHRS on October 14, 2020. An additional panel was sent to a new participant on March 30, 2021.

- *HTLV Test Kits*
  - Four different assays were used by the 16 participants (excluding the NLHRS) who returned results (Appendix 2).

- *Data entry*
  - Results entry for this panel utilized an in-house developed website.

Homogeneity and Stability

- The homogeneity and stability of the 2021Apr19 HTLV serology panel was assessed by comparing the participants' results (including the NLHRS) with the results of the panel's characterization performed by the NLHRS prior to the test event.
- There was no indication of heterogeneity or instability of the panel samples as the results submitted by the participants are consistent with the expected results from the NLHRS characterization of each panel member (Figure 1 and Appendix 1).

Results

- *Evaluation Criteria:*
  - Negative samples: HTLV non-reactive/negative in the final HTLV serology interpretation with assay results supporting the interpretation.
  - Positive samples: HTLV reactive/positive in the final HTLV serology interpretation with assay results supporting the interpretation. Participants must provide a recommendation for further action for samples that they could not determine the true serology status for based on the assay used in their testing.

- *Qualitative Group Analysis (Figure 1)*:
  - *Sample A (HTLV-II Ab Positive)* – 17/17 participants provided either a correct serology status and/or recommendation.
  - *Sample B (HTLV-I Ab Positive)* – 17/17 participants provided either a correct serology status and/or recommendation.
  - *Sample C (Negative)* – 17/17 participants provided either a correct serology status and/or recommendation.
  - *Sample D (HTLV-I Ab Positive)* – 17/17 participants provided either a correct serology status and/or recommendation.
  - *Sample E (Negative)* – 17/17 participants provided either a correct serology status and/or recommendation.

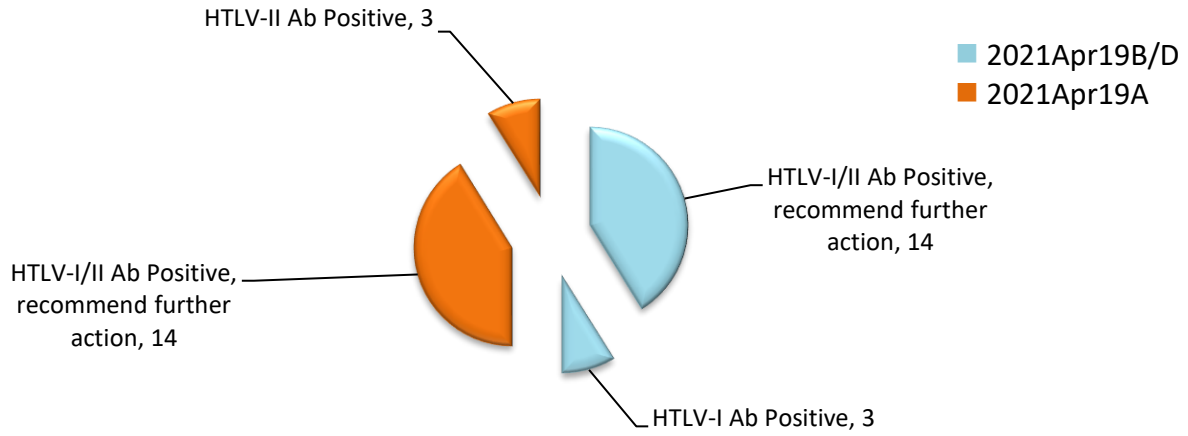

Figure 1: The final HTLV serology status of the positive samples in the 2021Apr19 HTLV serology panel submitted by participants using HTLV screening and confirmatory assays (including NLHRS).

Findings

All participants correctly identified the serology status and/or provided an appropriate recommendation for the panel samples included in the 2021Apr19 test event.

We value each laboratory's participation in these QA test events and your suggestions for improvement. The NLHRS is committed to improve all aspects of the HTLV serology proficiency testing program in order to provide quality proficiency testing to our participants.

If you have any comments, suggestions or concerns, please contact us at:

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Thank you for your participation in the NLHRS HTLV Serology Quality Assurance Program

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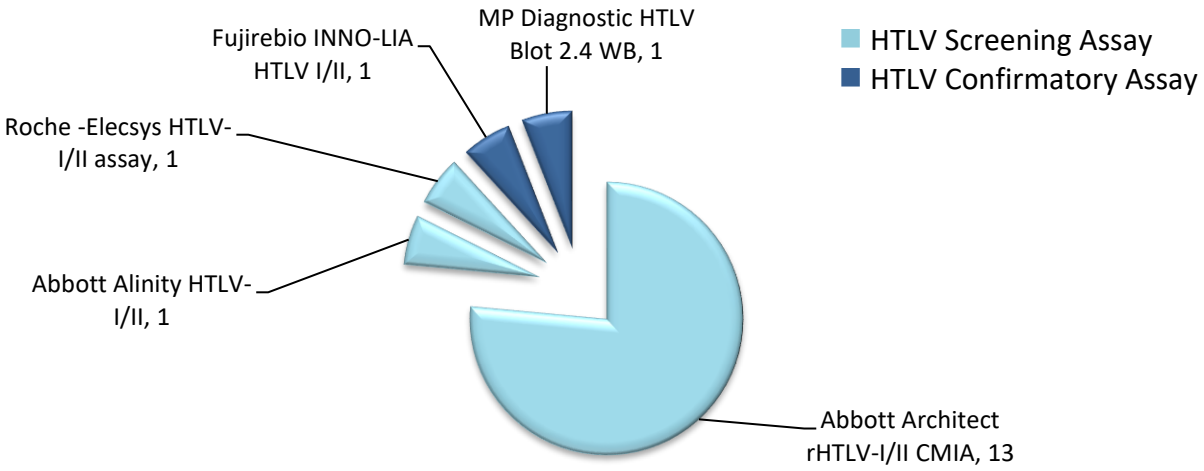
Appendix 1: NLHRS characterization of the 2021Apr19 HTLV serology panel samples

The NLHRS 2021Apr19 HTLV Panel Sample Testing Results									
Sample	Final Status	NLHRS Testing							
		Fujirebio INNO-LIA HTLV I/II Score							
		Interpretation	p19 I/II	p24 I/II	gp46 I/II	gp21 I/II	p19 I	gp46 I	gp46 II
A	HTLV-II Ab Positive	HTLV-II Positive	++	+++	+++	+++	-	-	+++
B	HTLV-I Ab Positive	HTLV-I Positive	+++	+++	+++	+++	++	+++	-
C	Negative	Negative	-	-	-	-	-	-	-
D	HTLV-I Ab Positive	HTLV-I Positive	+++	+++	+++	+++	++	+++	-
E	Negative	Negative	-	-	-	-	-	-	-

N/T: Not tested

Appendix 2: Summary of assays used by the participants in the 2021Apr19 HTLV test event



Appendix 3: Summary of bands detected in samples A, B, and D by the Fujirebio INNO-LIA HTLV-I/II and MP Diagnostic HTLV Blot 2.4 WB assays in the 2021Apr19 HTLV test event

Fujirebio INNO-LIA HTLV-I/II	Frequency of Bands Detected						
Sample	p19 I/II	p24 I/II	gp46 I/II	gp21 I/II	p19-I	gp46-I	gp46-II
2021Apr19A	2	2	2	2	1	1	2
2021Apr19B	2	2	2	2	2	2	-
2021Apr19D	2	2	2	2	2	2	-

MP Diagnostic HTLV Blot 2.4 WB	Frequency of Bands Detected										
Sample	rgp46-I	rgp46-II	p53	gp46	p36	p32	p28	P26	P24	P19	GD21
2021Apr19A	-	1	1	-	1	-	1	-	1	1	1
2021Apr19B	1	-	1	1	1	1	1	1	1	1	1
2021Apr19D	1	-	1	1	1	1	1	1	1	1	1

Appendix 4: Troubleshooting

Troubleshooting; common causes of outlying and/or aberrant results in serology and molecular Laboratories.

Type of Error	Possible Cause(s)	Pre-Analytical	Analytical	Post- Analytical
Sample mix-up	Can occur during specimen reception or testing. May result in outlying/aberrant results for one or all samples mixed-up.	✓	✓	
Transcription	• Incorrect test ordering by physician	✓		
	• Incorrect shipment address	✓		
	• Selecting the wrong assay for data entry	✓		
	• Interchanging results for two or more specimens			✓
	• Entering incorrect results			✓
	• Entering values in the incorrect field (e.g., OD as S/Co)			✓
	• Entering values in the incorrect unit (e.g., IU/mL instead of $\log_{10}$ copies/mL)			✓
	• Using a comma instead of a dot to denote a decimal point			✓
	• Selecting the incorrect assay interpretation or analyte			✓
	• Failure to recommend follow-up testing where necessary			✓
	It is recommended all results that are manually transcribed or entered electronically be checked by a second individual to avoid transcription errors.			
Outlying and/or Aberrant Results (random error)	Sporadic test results identified as outlying and/or aberrant can be classified as random events. Possible causes of random error include:			
	• Incorrect sample storage/shipping conditions	✓	✓	
	• Incorrect test method	✓	✓	
	• Insufficient mixing of sample, especially following freezing		✓	
	• Poor pipetting		✓	
	• Ineffective or inconsistent washing		✓	
	• Transcription errors	✓		✓
	• Cross-contamination or carryover	✓	✓	
	• Presence of inhibitors to PCR		✓	
Outlying and/or Aberrant Results (systematic error)	A series of test results identified as outlying and/or aberrant may be due to a systematic problem. Systematic problems may be due to:			
	• Reagents contaminated, expired, or subject to batch variation		✓	
	• Instrument error or malfunction		✓	
	• Insufficient washing		✓	
	• Incorrect wavelength used to read the assay result		✓	
	• Cycling times too long/short or temperature too high/low		✓	
	• Incubation time too long/short or temperature too high/low		✓	
	• Insufficient mixing/centrifuging before testing		✓	
	• Incorrect storage of test kits and/or reagents	✓		
	• Contamination of master-mix, extraction areas or equipment		✓	
	• Ineffective extraction process		✓	
	• Degradation of master-mix components		✓	
	• Suboptimal primer design (in-house assays)		✓	

This table was modified from a report produced by the National Reference Laboratory (NRL), Melbourne, Australia.